

Reduction of *Acinetobacter baumannii* biofilm formation by natural zeolite

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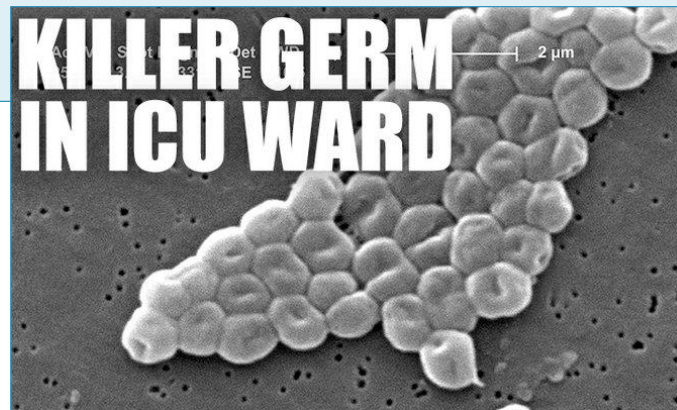
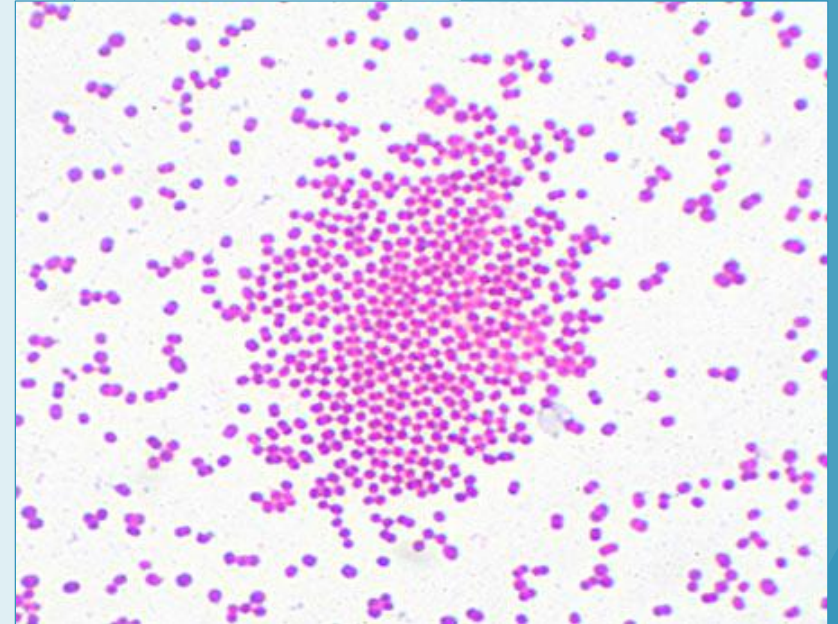
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Acinetobacter baumannii

- ▶ Gram negative coccobacillus
- ▶ Emerging human pathogen isolated mainly from hospital setting
- ▶ Pneumonia, bloodstream infections, urinary tract infections, wound infections in immunosuppressed patients
- ▶ Environmental isolates related to clinical isolates were found in soils and waters under the influence of human solid and liquid waste (Seine River, Sava River, acid paleosol from Croatia)
- ▶ Survives adverse environmental conditions for several months



Acinetobacter baumannii

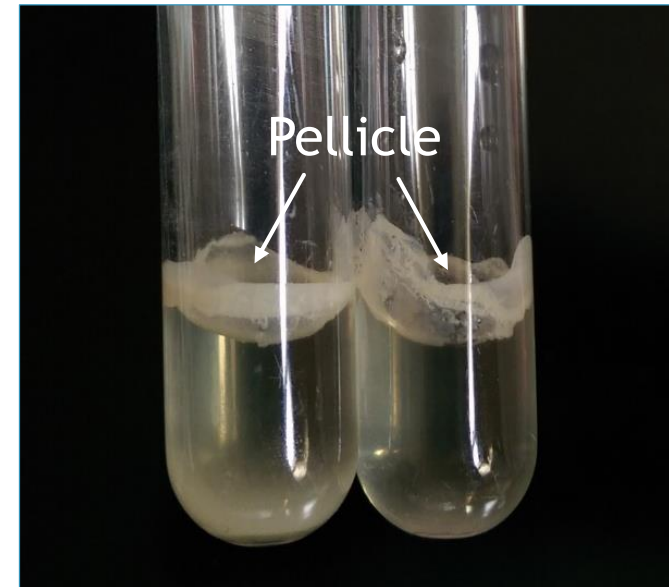
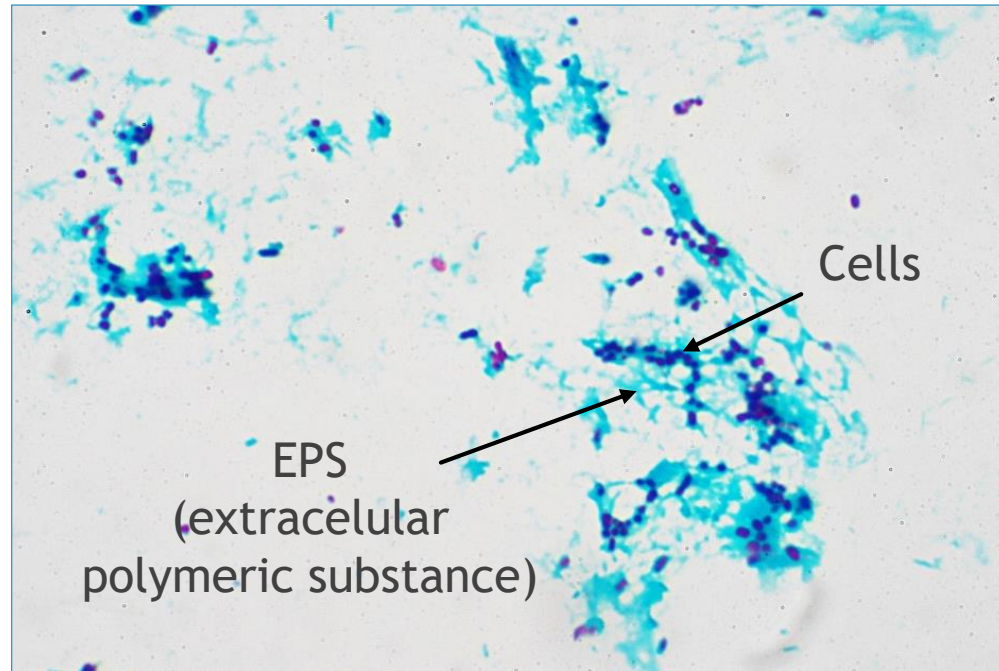
- ▶ Antibiotic resistance and virulence factors contribute to *A. baumannii* success as a pathogen
- ▶ Surface motility on solid/semi-solid media
- ▶ Biofilm formation on various surfaces
 - ▶ respiratory devices, intravenous devices, catheters, furniture, linen
 - ▶ human epithelial cells



Bacteria on human alveolar epithelial cells

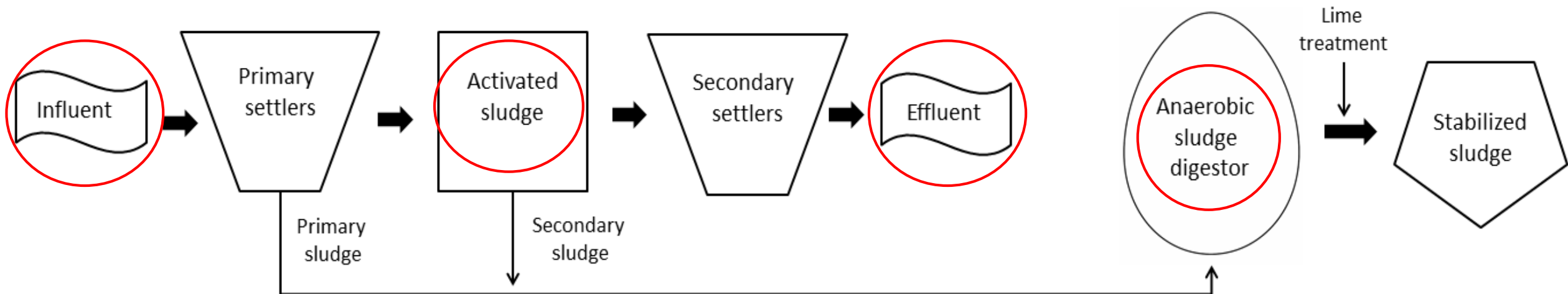
Acinetobacter baumannii

- ▶ Biofilm - an assemblage of cells enclosed in an extracellular matrix formed on different interfaces (solid-liquid, air-liquid)
- ▶ Pellicle - highly organised form of biofilm formed at the air-liquid interface



Experimental

- ▶ Wastewater treatment plant in Zagreb
- ▶ Combined sewage of domestic, hospital, industry and storm wastewater
- ▶ Samples of influent, effluent, fresh activated and digested sludge were analysed

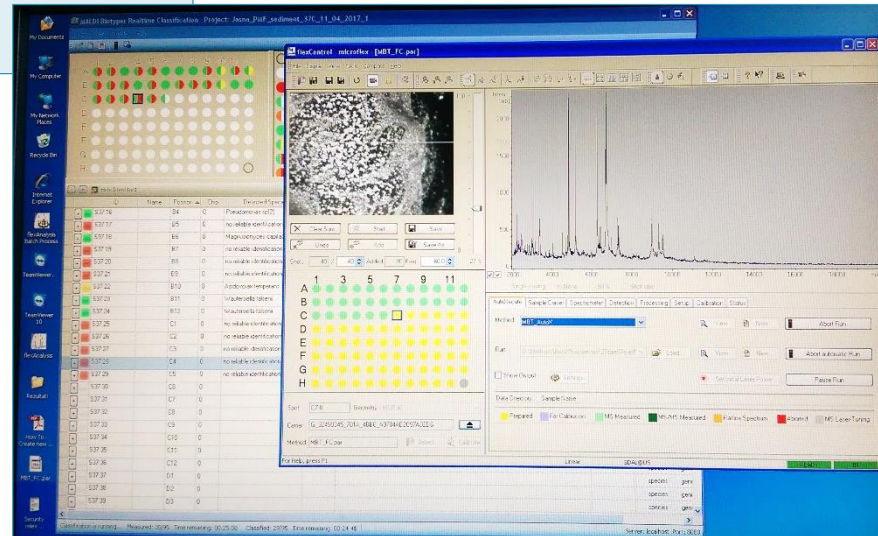


Experimental

- ▶ Isolation on commercial agar CHROMagar Acinetobacter at 42° C/48h
- ▶ Identification with Matrix assisted laser desorption ionisation with time of flight (MALDI TOF) on ribosomal proteins



CHROMagar Acinetobacter



MALDI TOF

Experimental

- ▶ Antibiotic resistance profile (Vitek2 system, EUCAST and CLSI criteria for clinical isolates)
- ▶ Hydrophobicity (BATH assay)- affinity of bacteria for organic hydrocarbon
- ▶ Biofilm formation (Crystal violet assay)
 - ▶ $OD_{550} < 0.3$ poor
 - ▶ $OD_{550} 0.3-1.0$ intermediate
 - ▶ $OD_{550} > 1.0$ strong
- ▶ Pellicle formation
 - ▶ No pellicle (0)
 - ▶ Poor (1)
 - ▶ Strong (2)



Vitek2



BATH assay



Crystal violet assay

Natural zeolite (NZ)

- ▶ Quarries at Donje Jesenje, Croatia
- ▶ Clinoptilolite (50-55%), celadonite, plagioclase feldspars and opal-CT (10-15% each), analcime and quartz in traces
- ▶ <math><0.122\text{mm}</math>
- ▶ dry NZ was sterilized by autoclaving
- ▶ Experiments with 1 and 10 wt % NZ



Selective immobilization of *Acinetobacter junii* on the natural zeolitized tuff in municipal wastewater

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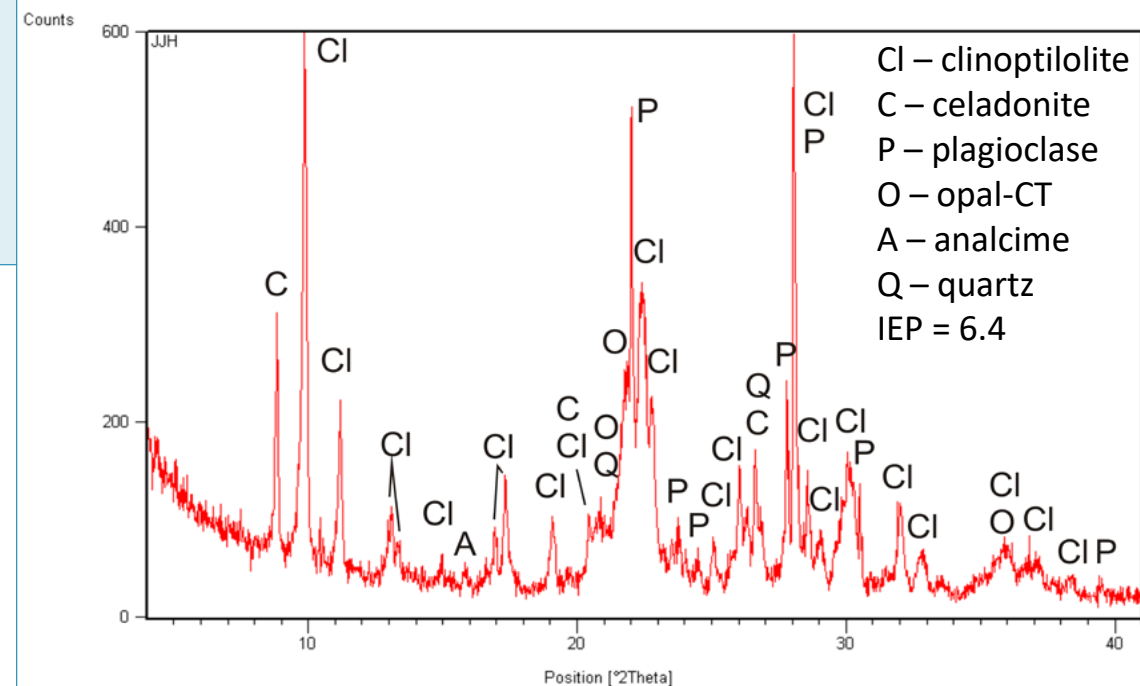
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ABSTRACT

The immobilization of desired bacteria onto material was usually performed in synthetic media. The aim of this study was to test the immobilization of phosphate (P)-accumulating bacteria *Acinetobacter junii* containing the common



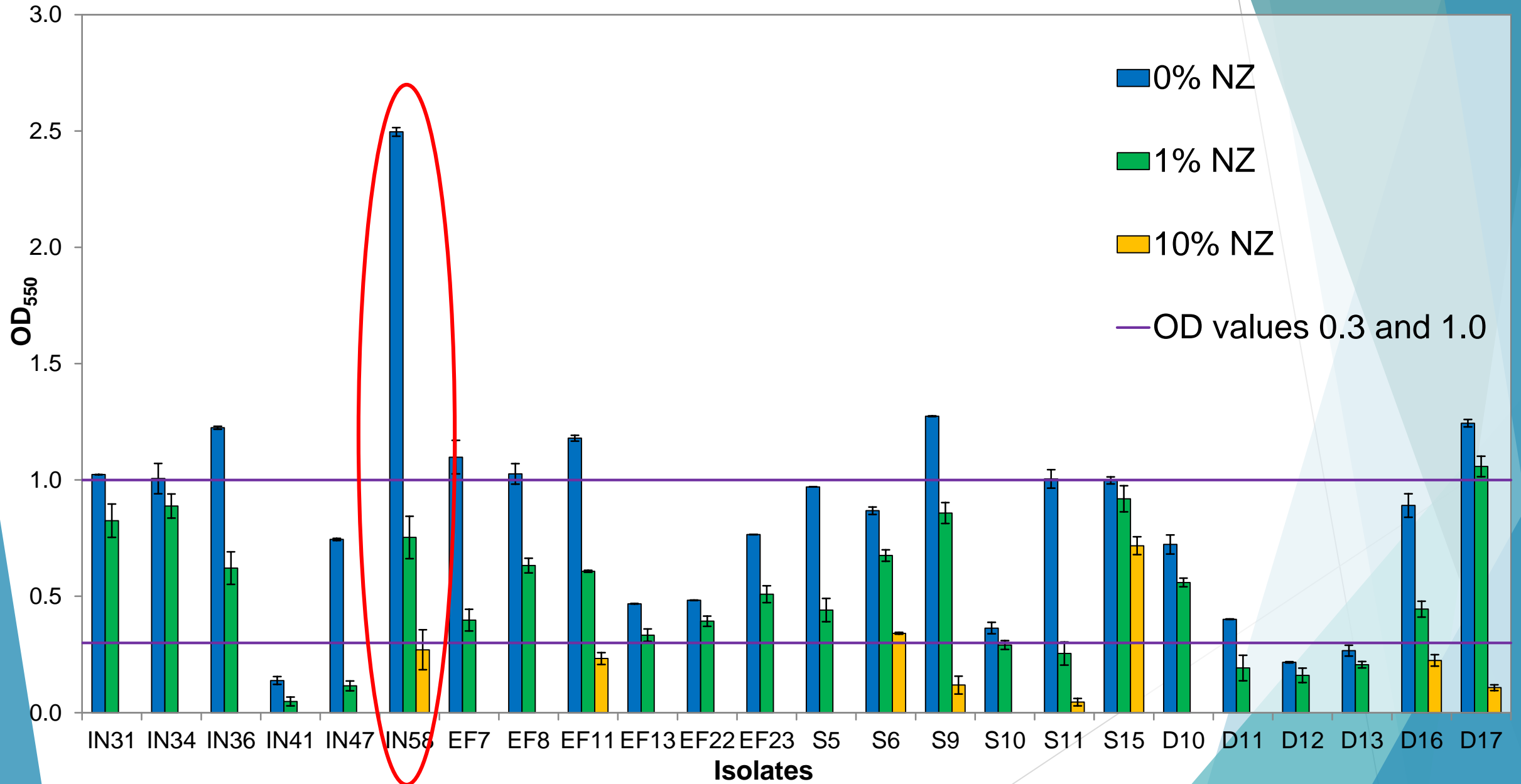
Isolate	Antibiotic resistance	Hydrophobicity (%)	Pellicle formation	Biofilm formation (OD ₅₅₀)
IN31	sensitive	97	1	1.024
IN34	MDR	1	1	1.006
IN36	sensitive	2	1	1.225
IN41	MDR	0	0	0.138
IN47	MDR	0	1	0.745
IN58	sensitive	93	2	2.497
EF7	MDR	0	1	1.098
EF8	MDR	0	1	1.026
EF11	sensitive	80	2	1.180
EF13	MDR	0	1	0.468
EF22	MDR	0	1	0.483
EF23	MDR	0	1	0.766
S5	MDR	3	1	0.971
S6	sensitive	78	1	0.868
S9	sensitive	8	2	1.274
S10	MDR	2	1	0.364
S11	MDR	0	1	1.005
S15	sensitive	79	1	0.998
D10	sensitive	0	1	0.723
D11	MDR	46	1	0.402
D12	MDR	49	1	0.217
D13	sensitive	0	1	0.267
D16	sensitive	67	1	0.891
D17	MDR	1	2	1.244

24 isolates recovered
14 MDR, 10 sensitive to 12 antibiotics tested
9/24 isolates hydrophobic
Majority of isolates intermediate biofilm and poor pellicle formers
Antibiotic sensitive isolates more hydrophobic and stronger biofilm and pellicle formers

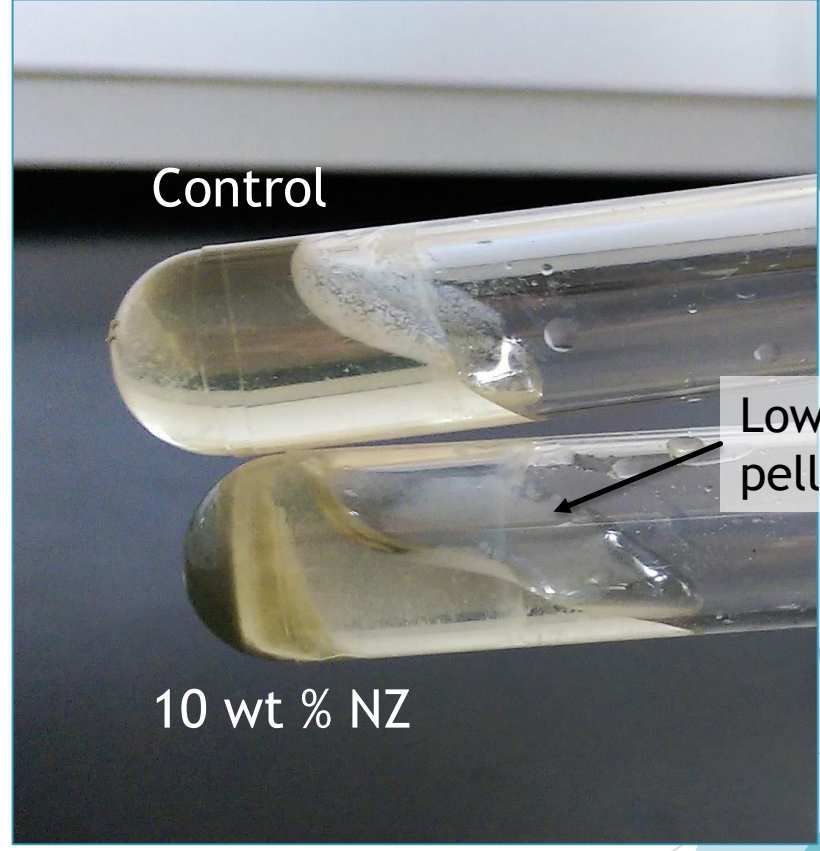
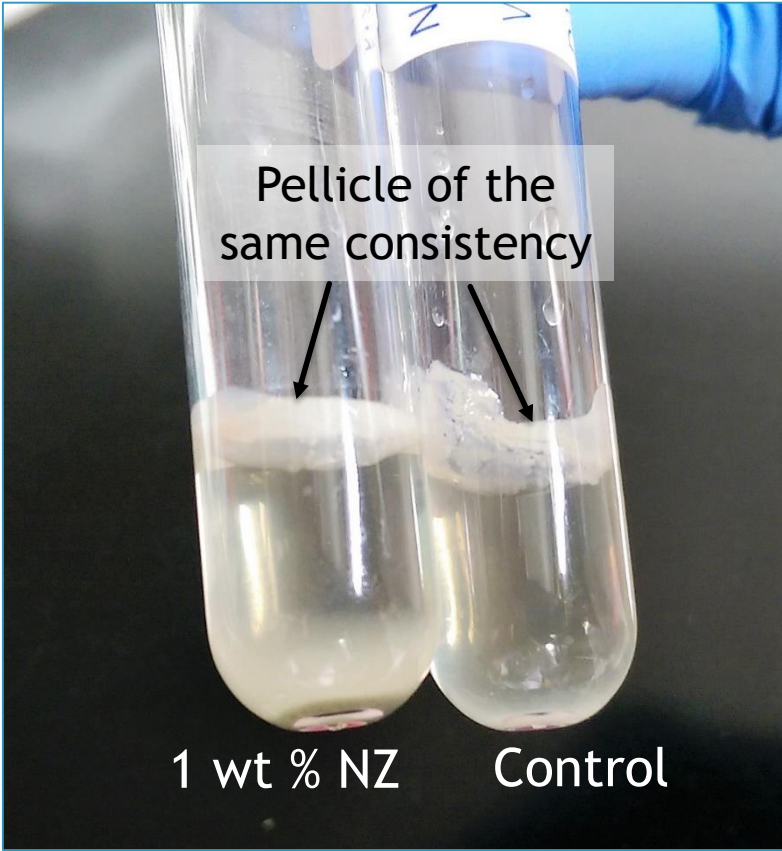
	Hydrophobicity	Biofilm	Pellicle
Hydrophobicity	1.000	r=0.425 p=0.003	r=0.433 p=0.002
Biofilm		1.000	r=0.682 p=0.000
Pellicle			1.000

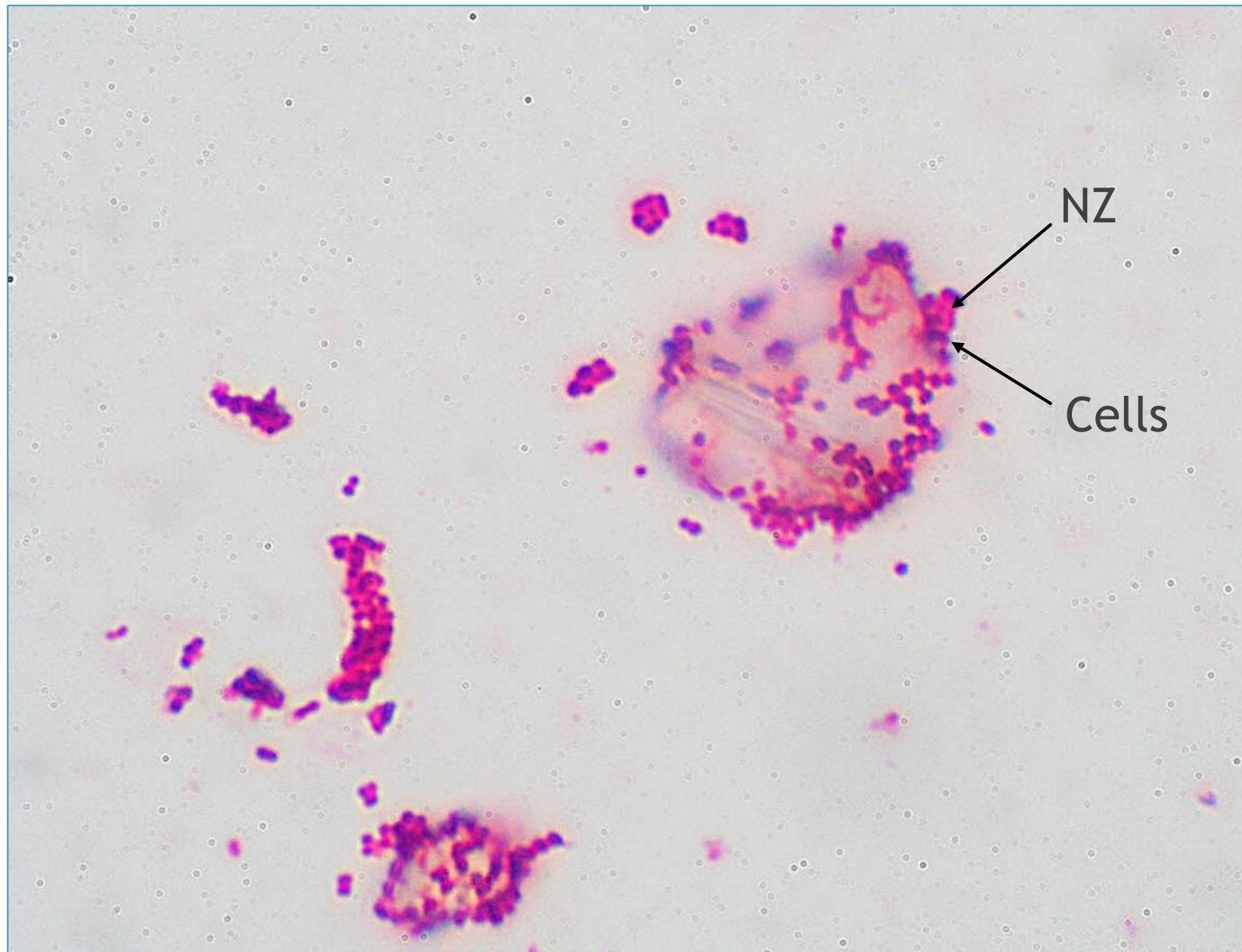
IN- influent, EF- effluent, S - fresh sludge, D - digested sludge
MDR (multi-drug resistant)- resistance to three or more classes of antibiotics

Biofilm formation



Pellicle formation





Cells of *Acinetobacter baumannii* immobilized onto NZ particles

Conclusions

- ▶ Cell surface hydrophobicity is an important feature which determines biofilm and pellicle formation of *A. baumannii*
- ▶ Isolates sensitive to antibiotics form stronger biofilm and pellicles than MDR isolates
- ▶ NZ successfully reduces biofilm and pellicle formation due to the immobilization of bacteria onto the NZ particles

Conclusions

- ▶ NZ is a promising material for the reduction of *A. baumannii* virulence factors
- ▶ NZ could find application in control of this emerging pathogen in the form of cleaning product where *A. baumannii* could be captured by NZ and safely removed from the contaminated environment

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THANK YOU!